

The role of the 5-lipoxygenase pathway in Alzheimer's disease

Molina Mhatre^{1,2}

¹Free Radical Biology and Aging Research Program, Oklahoma Medical Research Foundation, 825 NE 13th Street, Oklahoma City, OK 73104; ²Department of Psychiatry and Behavioral Sciences, University of Oklahoma Health Science Center, Oklahoma City, OK 73104, USA. Correspondence: e-mail: Molina-Mhatre@omrf.ouhsc.edu

CONTENTS

Abstract	83
Introduction	83
Neuroinflammation and age-related neurodegenerative diseases	83
Neuroinflammation as a cause or a consequence of amyloid pathology	84
Chronic NSAID treatment reduces the risk of AD	84
Arachidonic acid metabolism	85
Natural polyphenols that act as 5-LOX inhibitors	85
5-LOX and AD	86
Neuroprotective actions of 5-LOX inhibitors	87
Modulation of signal transduction by 5-LOX inhibitors	87
Conclusions	87
References	87

Abstract

Many neurological diseases are now recognized to involve neuroinflammation as a major pathological feature. Neuroinflammation can be both a cause and a consequence of the disease process in Alzheimer's disease (AD). Chronic use of nonsteroidal antiinflammatory drugs (NSAIDs) has been found to delay the onset of AD and slow disease progression, a finding that has stimulated substantial interest in the role of these drugs in the prevention and treatment of AD. Unfortunately, recent clinical trials involving NSAIDs either have produced negative results or have been suspended due to toxicity issues concerning cyclooxygenase inhibition (e.g., ADAPT). Consequently, there is an urgent need to develop and test new antiinflammatory drugs devoid of the toxic effects associated with cyclooxygenase inhibition but which still suppress neuroinflammation and β -amyloid-induced neurotoxicity. In addition to the cyclooxygenase pathway, arachidonic acid is also metabolized by the lipoxygenase pathway to form leukotrienes and lipoxins. There is some evidence indicating that 5-lipoxygenase inhibitors may offer protection against various aspects of the pathogenesis of AD. Based on these findings, we suggest that 5-lipoxygenase inhibitors may have therapeutic potential in the prevention and treatment of AD.

Introduction

The brain of patients with Alzheimer's disease (AD) shows chronic inflammatory responses characterized by activated microglia and astrocytes and enhanced expression of cytokines and complement factors surrounding β -amyloid (A β) deposits. Several epidemiological studies have demonstrated a lower risk for AD in chronic users of nonsteroidal antiinflammatory drugs (NSAIDs). This finding has stimulated substantial interest in the role of these drugs in the prevention and treatment of AD. Unfortunately, recent clinical trials on the use of these drugs have produced negative results or have been suspended due to toxicity associated with cyclooxygenase inhibition. Therefore, there is an urgent need to develop and test new antiinflammatory drugs without the toxic effects associated with cyclooxygenase inhibition but retaining the ability to suppress neuroinflammation. This review article concerns the potential use of 5-lipoxygenase (5-LOX) inhibitors for the prevention and treatment of AD.

Neuroinflammation and age-related neurodegenerative diseases

Neuroinflammation is widely considered as a possible pathophysiological mechanism of aging-associated neurodegeneration. It is thought to be an integral component of normal aging, as well as in the pathogenesis of age-related neurological diseases such as AD, Parkinson's disease and amyotrophic lateral sclerosis (ALS). These neurodegenerative diseases share neuroinflammatory characteristics such as elevation of proinflammatory cytokines (i.e., IL-1 and TNF- α), microglial activation and the presence of reactive astrocytes. Numerous research reports have provided evidence of neuroinflammation in the brain of AD patients (e.g., 1-4). Extracellular deposition of A β , a pathological characteristic of AD, has been reported to trigger many signal transduction pathways that contribute to neuroinflammation. Amyloid deposits are found to be co-localized with various inflammation-related proteins, such as complement proteins, acute-

phase proteins and clusters of activated microglia, indicating a strong glial response to amyloid deposits. Elevated levels of cytokines, cytokine receptors and reactive astrocytes suggesting inflammation have also been reported in post mortem AD brains (5-7).

Neuroinflammation as a cause or a consequence of amyloid pathology

Although the extent of involvement of neuroinflammation in AD pathogenesis is still unclear, it appears to be both a consequence and a cause of amyloid pathology. The theory that chronic inflammation may accelerate AD pathogenesis is supported by recent genetic findings showing that polymorphisms in proinflammatory genes (e.g., IL-1 α , IL-1 β and TNF- α) enhance the risk for AD (8, 9). The extensive neuronal damage observed in AD patients is also thought to be a consequence of glial activation and inflammation, since glial activation and cytokine overexpression are observed decades before the pathological and behavioral changes selective for AD occur in individuals with Down's syndrome and traumatic brain injury (10).

Activated microglia and reactive astrocytes in AD brains were initially considered to be a response to amyloid deposition. However, there is a consensus that chronic microglial activation may also accelerate the amyloid cascade by stimulating the secretion of proinflammatory cytokines and reactive molecules that further enhance inflammation (10). This is supported by observations whereby microglia, when stimulated *in vitro*, generate free radicals, proinflammatory cytokines and neurotoxic lipid mediators. Proinflammatory cytokines may further be involved in activating β -secretase and stimulating more amyloid production. Enhancement of amyloid deposition has been observed in APPV717F transgenic mice upon administration of bacterial lipopolysaccharide (LPS). Chronic glial activation was shown to enhance the hyperphosphorylation of tau, resulting in the subsequent development of neurofibrillary tangles, an important pathological feature of AD (11). These observations confirm that neuroinflammation may accelerate amyloid deposition and the formation of neurofibrillary tangles in a high-risk group for AD (12, 13).

Based on this link between neuroinflammation and AD pathology, it appears logical that suppression of microglial activation and free radical generation should delay the progression of the disease. Therefore, numerous research laboratories have focused their efforts on antiinflammatory drugs and their effects on various aspects involved in the pathogenesis of AD.

Chronic NSAID treatment reduces the risk of AD

The first data associating chronic neuroinflammation with the pathogenesis of AD came from epidemiological studies. These studies showed a reduction of up to 50% in the risk of AD and lower rates of cognitive decline in chronic NSAID users (14-16), and subsequent studies by

various groups confirmed these findings. The results of these epidemiological observations were further supported by observations from experimental studies using animal models of AD. Chronic treatment with a subset of NSAIDs (e.g., ibuprofen, flurbiprofen, indomethacin) reduced neuroinflammation, A β levels and the deposition of A β in rat brain (17-23). In a study by Lim *et al.* (22), 6 months of oral treatment with ibuprofen decreased amyloid plaque number, as well as SDS-soluble and -insoluble A β levels in transgenic Tg2576 mice. Ibuprofen treatment suppressed the number of activated microglia and brain levels of IL-1 β , a key cytokine implicated in AD pathogenesis (17, 21, 22). These observations were confirmed by Yan and coworkers (23).

However, the antiinflammatory mechanism underlying the beneficial effects of NSAIDs on AD pathogenesis was challenged by recent findings demonstrating selective suppression of A β 42 generation by certain NSAIDs, such as ibuprofen, indomethacin and sulindac sulfide (24). These compounds were suggested to change the conformation of the γ -secretase complex by binding to a novel site distinct from the catalytic center. However, further support for the antiinflammatory mechanism of NSAIDs in protecting against AD comes from the finding of their ability to suppress various markers of inflammation, which are considered to be important in animal models of AD (25, 26). These studies have shown that NSAIDs may exert their beneficial effects via multiple mechanisms of action (reviewed in 27).

Based on the epidemiological studies suggesting a protective role for NSAIDs, these drugs were expected to be excellent drug candidates for the treatment and/or prevention of AD. However, the results of clinical trials in AD patients have been quite discouraging. Also, the low tolerability of these drugs due to gastric and renal sensitivity and liver damage in elderly patients has clouded the results from these trials. The dropout rates from some of these clinical trials were as high as 50%.

At present, there is substantial evidence that both cyclooxygenase COX-1 and COX-2 isoforms are not only involved in homeostasis but are also modulators of inflammatory reactions. Recent observations indicate that treatment with COX-2-selective NSAIDs, such as rofecoxib, is associated with increased cardiovascular risk. This compound was withdrawn from the market by Merck & Co. after a colon cancer prevention trial revealed that it was associated with double the rate of strokes and heart attacks compared to placebo. It was thought that selective targeting of COX-2 could lead to cardiovascular effects by altering the fine balance between the fatty acids prostacyclin and thromboxane, which control blood clotting. In a study conducted by Pfizer, patients with cardiac surgery who were taking both aspirin and COX-2 inhibitors reported nearly 3 times the rate of cardiovascular events compared with those on placebo, indicating that inhibition of COX-2 increases the risk for cardiovascular events. This implies that the use of NSAIDs, which inhibit both COX-1 and COX-2, might also increase the risk for cardiovascular events.

The concern for toxicity of NSAIDs was reflected in the announcement by the National Institutes of Health that the use of naproxen (nonselective COX inhibitor) and celecoxib (selective COX-2 inhibitor) was being suspended in a large AD prevention trial. The trial, called the Alzheimer's Disease Anti-Inflammatory Prevention Trial (or ADAPT), was designed to assess the potential benefit of long-term use of NSAIDs in decreasing the risk of developing AD in people 70 years of age or older, who were considered to be at an increased risk for the disease. Even though the mechanism implicated in lowering toxic A β production by NSAIDs is considered not to involve COX inhibition, all NSAIDs reported to reduce A β 42 also potentially inhibit COX activity at much lower doses and are therefore likely to have toxicity problems. The (*R*)-enantiomers in the profen NSAIDs supposedly lack COX-inhibitory activity but retain the ability to reduce A β 42 production. These (*R*)-enantiomers are currently the focus of research by many investigators (e.g., 28, 29).

In conclusion, the efficacy of NSAIDs in AD has not yet been proven and safety issues have significantly limited their use. Consequently, there is an urgent need to develop and test new antiinflammatory drugs without the toxic effects associated with COX inhibition but retaining the ability to suppress neuroinflammation and toxicity induced by A β 42.

Arachidonic acid metabolism

Arachidonic acid and lysophospholipids are released by the action of phospholipase A₂ (PLA₂) on phospholipids. In mammalian brain, there are different forms of phospholipases and they are implicated in inflammation, neurodegeneration and the intracellular as well as intercellular signal transduction network. Elevated immunoreactivity for PLA₂ has been observed in association with amyloid deposits in AD brain. A β peptides have also been observed to activate this enzyme in *in vitro* studies. In mammalian cells, the release of arachidonic acid constitutes the rate-limiting step in the biosynthesis of eicosanoids, such as prostaglandins, leukotrienes, thromboxanes and platelet-activating factor, all of which act as potent inflammatory mediators. High levels of these metabolites are neurotoxic and are associated with neurodegeneration. NSAIDs block the production of prostaglandins by inhibiting COX-mediated metabolism of arachidonic acid.

In addition to the cyclooxygenase pathway, arachidonic acid is also metabolized by the lipoxygenase pathway (Fig. 1) to form leukotrienes and lipoxins. There are 6 mammalian lipoxygenases that site-specifically oxidize arachidonic acid to lipid hydroperoxides and are classified as 5-, 8-, 12- and 15-lipoxygenases according to the carbon atom of arachidonic acid at which oxygen is inserted. The action of 12-lipoxygenase leads to the formation of oxidized lipids such as 12(*S*)-hydroxyeicosatetraenoic acid [12(*S*)-HETE] (30, 31).

The major lipoxygenase of the central nervous system is 5-lipoxygenase (5-LOX), which is present in neurons

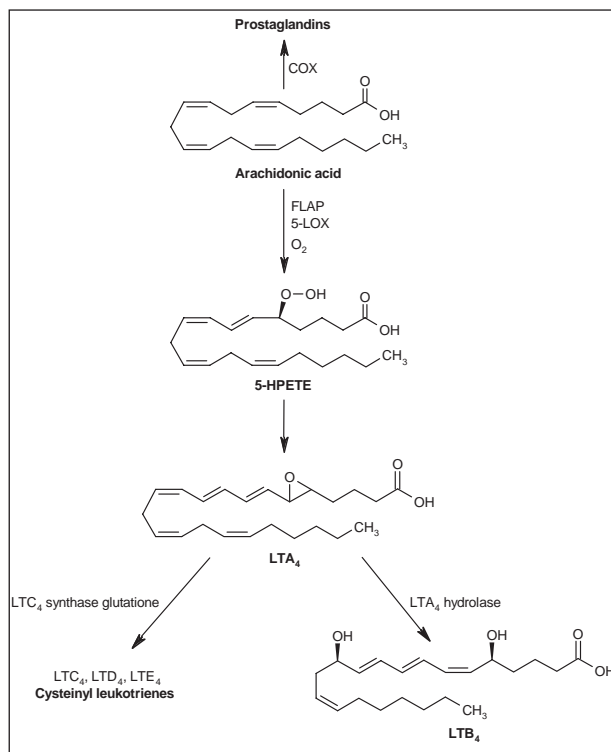


Fig. 1. The 5-lipoxygenase pathway of arachidonic acid metabolism.

and glial cells at high levels, and its expression increases in the CNS as a function of age (32, 33). 5-LOX catalyzes the first step in a pathway leading to leukotriene end products (33-36). When unstimulated, 5-LOX resides as a cytosolic enzyme which is activated by p38 mitogen-activated protein kinase and gets translocated to the nucleus (34, 38). In the nucleus, the interaction between 5-LOX and the substrate arachidonic acid is mediated by the 5-LOX-associated protein, FLAP (34, 38). Thus, 5-LOX activity appears to be regulated at multiple levels, both pre- and posttranscriptional.

The first product of 5-LOX catalysis is leukotriene A₄ (LTA₄), which can be hydrolyzed to the dihydroxy lipid product leukotriene B₄ (LTB₄) or, alternatively, LTA₄ can be conjugated with glutathione and further processed to yield a series of cysteinyl leukotrienes (LTC₄, LTD₄ and LTE₄; Fig. 1). Leukotrienes can be released from the cell as paracrine agents to promote bronchoconstriction, platelet aggregation or other tissue-specific functions (34-37). Accordingly, leukotriene antagonists are being investigated for the treatment for asthma but have not been investigated for their role in the pathogenesis of neurodegenerative disorders.

Natural polyphenols that act as 5-LOX inhibitors

The two classical natural products that inhibit 5-LOX are nordihydroguaiaretic acid (NDGA) and the curry spice component curcumin (Fig. 2). NDGA inhibits 5-LOX, 12-LOX and 15-LOX with reported K_i values of 200 nM, 30

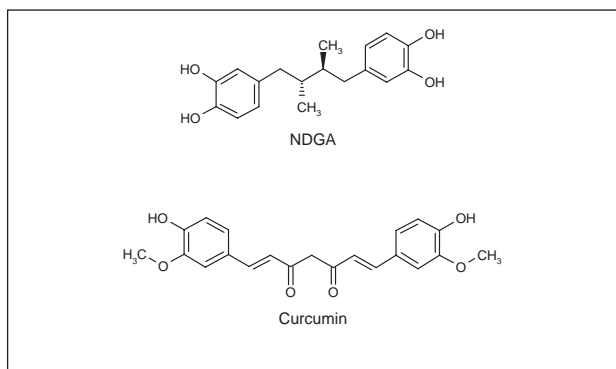


Fig. 2. Structures of the 5-lipoxygenase antagonists NDGA and curcumin.

μM and $30 \mu\text{M}$, respectively (39). The mechanism underlying NDGA-induced 5-LOX inhibition is thought to involve reduction of the iron center in the enzyme by the catechol group of the compound (35, 40). Similar to NDGA, curcumin (turmeric) also has a polyphenolic structure (Fig. 2). Curcumin also inhibits 5-LOX, but less efficiently than NDGA (41). Interestingly, recent work from Cole's laboratory shows that curcumin reduces amyloid deposition and IL-1 β expression in a transgenic model of AD amyloidopathy (42). Oral curcumin also reduces loss in synaptic markers following intracerebroventricular treatment with A β .

While considerable importance has been given to cyclooxygenase as an inflammatory contributor to neurological diseases (reviewed in 43), partially due to the protective benefits against AD that have been associated with chronic use of NSAIDs (1), much less attention has been given to the lipoxygenase branch of arachidonate metabolism. However, recent data from the literature and some data from our laboratory suggest that lipoxygenase inhibitors are excellent drug candidates for clinical development for the treatment of neurodegenerative diseases (reviewed in 44). In support of this argument, we have recently found that dietary NDGA delays neurodegeneration in a mouse model of amyotrophic lateral sclerosis, indicating a broader role of the lipoxygenase pathway in causing neuronal degeneration (45).

There is evidence implying that the 5-LOX pathway may be involved in AD pathogenesis. Frautschy *et al.* (46) have shown that dietary curcumin (a 5-LOX inhibitor and antioxidant) treatment prevents A β -induced spatial memory deficits and reduces A β deposits and microgliosis. Dietary curcumin but not ibuprofen also reduced oxidative damage and the loss of synaptophysin in this mouse model of AD.

Since A β accumulation is a pathological characteristic of AD, inhibition of the accumulation of A β peptide and the formation of A β fibrils from A β , as well as the destabilization of preformed A β fibrils in the CNS, would be attractive therapeutic targets for the treatment of AD. Natural polyphenols like curcumin are much more potent in inhibiting A β aggregation than ibuprofen and naproxen. Ono *et al.* (47, 48) reported that both curcumin and NDGA

concentration-dependently ($5\text{--}30 \mu\text{M}$) inhibited A β fibril formation from A β (1-40) and A β (1-42). These compounds also dose-dependently destabilized preformed A β fibrils. Soluble A β oligomers are more diffusible and more toxic and increasingly viewed as playing an important role in AD pathogenesis. Low micromolar and even submicromolar concentrations of curcumin effectively block soluble A β oligomer formation and toxicity. In a report by Ono *et al.*, NDGA was also shown to disaggregate A β protofibrils (47), whereas observations by Moss *et al.* indicated inhibition of direct protofibril-protofibril association of A β by NDGA (49). It is possible that the anti-amyloidogenic activity of NDGA and curcumin is 5-LOX-independent and may be due to their polyphenolic nature, as well the propensity of these compounds to bind to specific sites of A β or their metal-binding property (47, 48). The effects of NDGA and curcumin on A β aggregation could result in reducing the toxicity of A β . Consistent with this, Goodman *et al.* (50) previously reported that NDGA concentration-dependently reduces the cytotoxicity of A β to cultured rat hippocampal neurons by suppressing A β -induced accumulation of reactive oxygen species and intracellular free Ca $^{2+}$. These findings suggest that NDGA interferes with a neurodegenerative pathway important to the pathophysiology of AD.

5-LOX and AD

Naturally occurring polymorphisms in the 5-LOX promoter may influence expression levels of this protein in humans (51). Approximately 25% of the human population has a mutation in the 5-LOX promoter, which diminishes the expression of the 5-LOX gene. Manev and coworkers (52) hypothesized that 5-LOX promoter polymorphism could affect the onset of AD and/or influence the response of AD patients to anti-inflammatory treatment with 5-LOX inhibitors. The epidemiological implications of such promoter variation have not yet been rigorously investigated.

We have some evidence that 5-LOX levels are dysregulated in AD patients and in transgenic animal models of ALS and AD. Preliminary Western blot analysis of AD and age-matched brains of individuals who did not suffer from AD indicate highly variable expression of 5-LOX in AD brain cortex. However, the average level of 5-LOX was 2.8-fold greater in AD cortex than in normal cortex. Similarly, 5-LOX levels are increased in the cortex of the APP/PS1 mouse model of AD. 5-LOX mRNA was increased by at least 2-fold at 120 days in the spinal cord of transgenic G93A mice (a model of ALS) relative to the levels in nontransgenic mice, suggesting that changes in 5-LOX expression may be a factor associated with neurodegeneration.

Other studies have reported that in addition to 5-LOX, 12/15-LOX may also be important in AD pathogenesis. Studies have shown the presence of 12/15-LOX in the brain (53), and recently, levels of the 12/15-LOX metabolite (12/15-HETE) were found to be markedly elevated in AD brains compared to controls. The increase in the lev-

els of this metabolite was also directly correlated with brain lipid peroxidation (54).

Neuroprotective actions of 5-LOX inhibitors

Woo and coworkers have shown previously that another 5-LOX metabolite, LTB₄, mediates the release of free radicals from cultured fibroblasts exposed to TNF- α (55). EOC-20 cells stimulated with TNF- α produce robust amounts of reactive oxygen species as evidenced by nitrite accumulation in the medium. Inhibitors of arachidonic acid metabolism, especially 5-LOX inhibitors, are potent antagonists of TNF- α -induced nitrite production (56). TNF- α has been reported to play an important role in AD neurodegeneration (reviewed in 57). There are also reports on the involvement of nitric oxide in AD pathology (58, 59) and elevated levels of nitrotyrosine-modified proteins have also been found in AD brains (59-61). Thus, a potent inhibitory role of NDGA in suppressing TNF- α -induced nitrite production in glial cells (56) could be an important mechanism of NDGA's protective effect against AD pathogenesis.

Modulation of signal transduction by 5-LOX inhibitors

Recent data from several laboratories suggest that leukotrienes, particularly LTB₄, may modulate signal transduction. For instance, Funk and colleagues suggest that nuclear LTB₄ binds specific transcription factors including PPAR γ and/or AP1 (62), increasing the potency of gene induction. 5-LOX inhibitors have been reported to suppress the nuclear factor NF- κ B activation pathway (63). The 5-LOX inhibitor curcumin is known to directly inhibit I κ B kinase activity and I κ B α phosphorylation and subsequently block NF- κ B activation (64). There is evidence that NF- κ B is activated by A β as well as free radicals and inflammatory stimuli such as TNF- α and IL-1, etc. (Mhatre, unpublished observations). In human neuron-like cells, binding of A β to p75NTR (a member of the cell death receptor family) was found to activate NF- κ B and induce DNA fragmentation (65). Inhibition of NF- κ B activation by curcumin prevents A β -induced cell death, thus implicating NF- κ B in A β -induced cell death (66).

Conclusions

The use of anticholinesterase drugs and NSAIDs in the treatment of AD is limited by serious toxicity issues and contraindications such as gastric sensitivity, which is common in elderly patients. There is considerable hope that inhibition of neuroinflammatory processes might lead to a treatment approach to delay the onset of AD in individuals at high risk. Research from our laboratory and others suggests that some natural product inhibitors of 5-LOX might provide lead compounds for such pharmacotherapeutic development.

The 5-LOX inhibitors curcumin and NDGA possess both antioxidant and antiinflammatory activity. 5-LOX

inhibitors have been reported to block the increase in APP secretion by IL-1 β (66) and reduce A β aggregation (46, 47), which is considered critical for A β -induced neurotoxicity. 5-LOX levels are increased in neurons during aging and this could significantly increase the brain's vulnerability to neurodegeneration. As mentioned above, we have found that NDGA delays neurodegeneration in a mouse model of amyotrophic lateral sclerosis, implying a broader role of the LOX pathway in neuronal degeneration (45). Thus, based on these observations, we propose that the inhibition of arachidonic acid-5-LOX has therapeutic potential in the treatment and prevention of AD.

References

1. Eikelenboom, P., Veerhuis, R. *The role of complement and activated microglia in the pathogenesis of Alzheimer's disease*. Neurobiol Aging 1996, 17: 673-80.
2. McGeer, P.L., Akiyama, H., Itagaki, S., McGeer, E.G. *Activation of the classical complement pathway in brain tissue of Alzheimer patients*. Neurosci Lett 1989, 107: 341-6.
3. McGeer, P.L., Akiyama, H., Itagaki, S., McGeer, E.G. *Immune system response in Alzheimer's disease*. Can J Neurol Sci 1989, 16: 516-27.
4. McGeer, P.L., Itagaki, S., Tago, H., McGeer, E.G. *Reactive microglia in patients with senile dementia of the Alzheimer type are positive for the histocompatibility glycoprotein HLA-DR*. Neurosci Lett 1987, 79: 195-200.
5. Fillit, H., Ding, W.H., Buee, L. *Elevated circulating TNF levels in Alzheimer's disease*. Neurosci Lett 1991, 129: 318-20.
6. Griffin, W.S., Stanley, L.C., Ling, C. et al. *Brain interleukin 1 and S-100 immunoreactivity are elevated in Down syndrome and Alzheimer disease*. Proc Natl Acad Sci USA 1989, 86: 7611-5.
7. Griffin, W.S., Sheng, J.G., Roberts, G.W., Mrak, R.E. *Interleukin-1 expression in different plaque types in Alzheimer's disease: Significance in plaque evolution*. J Neuropathol Exp Neurol 1995, 54: 276-81.
8. Papassotiropoulos, A., Bagli, M., Jessen, F., Bayer, T.A., Maier, W., Rao, M.L., Heun, R. *A genetic variation of the inflammatory cytokine interleukin-6 delays the initial onset and reduces the risk for sporadic Alzheimer's disease*. Ann Neurol 1999, 45: 666-8.
9. McCusker, S.M., Curran, M.D., Dynan, K.B. et al. *Association between polymorphism in regulatory region of gene encoding tumour necrosis factor alpha and risk of Alzheimer's disease and vascular dementia: a case-control study*. Lancet 2001, 357: 436-9.
10. Mrak, R.E., Griffin, W.S. *Glia and their cytokines in progression of neurodegeneration*. Neurobiol Aging 2005, 26: 349-54.
11. Kitazawa, M., Yamasaki, T.R., Laferla, F.M. *Microglia as a potential bridge between the amyloid beta-peptide and tau*. Ann NY Acad Sci 2004, 1035: 85-103.
12. Guo, J.T., Yu, J., Grass, D., de Beer, F.C., Kindy, M.S. *Inflammation-dependent cerebral deposition of serum amyloid A protein in a mouse model of amyloidosis*. J Neurosci 2002, 22: 5900-9.

13. Qiao, X., Cummins, D.J., Paul, S.M. *Neuroinflammation-induced acceleration of amyloid deposition in the APPV717F transgenic mouse*. Eur J Neurosci 2001, 14: 474-82.
14. Mackenzie, I.R., Munoz, D.G. *Nonsteroidal anti-inflammatory drug use and Alzheimer-type pathology in aging*. Neurology 1998, 50: 986-90.
15. McGeer, P.L., McGeer, E.G., Rogers, J., Sibley, J. *Antiinflammatory drugs and Alzheimer's disease*. Lancet 1990, 335: 1037-9.
16. Rich, J.B., Rasmusson, D.X., Folstein, M.F., Carson, K.A., Kawas, C., Brandt, J. *Nonsteroidal anti-inflammatory drugs in Alzheimer's disease*. Neurology 1995, 45: 51-5.
17. Lim, G.P., Yang, F., Chu, T. et al. *Ibuprofen effects on Alzheimer pathology and open field activity in APPsw transgenic mice*. Neurobiol Aging 2001, 22: 983-91.
18. Cole, G.M., Morihara, T., Lim, G.P., Yang, F., Begum, A., Frautschy, S.A. *NSAIDs and antioxidant prevention of Alzheimer's disease: Lessons from in vitro and animal models*. Ann NY Acad Sci 2004, 1035: 68-84.
19. Sung, S., Yang, H., Uryu, K. et al. *Modulation of nuclear factor- κ B activity by indomethacin influences A β levels but not A β precursor protein metabolism in a model of Alzheimer's disease*. Am J Pathol 2004, 165: 2197-206.
20. Giovannini, M.G., Scali, C., Prosperi, C., Bellucci, A., Pepeu, G., Casamenti, F. *Experimental brain inflammation and neurodegeneration as model of Alzheimer's disease: Protective effects of selective COX-2 inhibitors*. Int J Immunopathol Pharmacol 2003, 16(2, Suppl.): 31-40.
21. Richardson, R.L., Kim, E.M., Shephard, R.A., Gardiner, T., Cleary, J., O'Hare, E. *Behavioural and histopathological analyses of ibuprofen treatment on the effect of aggregated A β (1-42) injections in the rat*. Brain Res 2002, 954: 1-10.
22. Lim, G.P., Yang, F., Chu, T. et al. *Ibuprofen suppresses plaque pathology and inflammation in a mouse model for Alzheimer's disease*. J Neurosci 2000, 20: 5709-14.
23. Yan, Q., Zhang, J., Liu, H. et al. *Anti-inflammatory drug therapy alters β -amyloid processing and deposition in an animal model of Alzheimer's disease*. J Neurosci 2003, 23: 7504-9.
24. Weggen, S., Eriksen, J.L., Das, P. et al. *A subset of NSAIDs lowers amyloidogenic A β 42 independently of cyclooxygenase activity*. Nature 2001, 414: 212-6.
25. Morihara, T., Teter, B., Yang, F. et al. *Ibuprofen suppresses interleukin-1 β induction of pro-amyloidogenic α (1)-antichymotrypsin to ameliorate beta-amyloid (A β) pathology in Alzheimer's models*. Neuropsychopharmacology 2005, 30: 1111-20.
26. Frautschy, S., Hu, W., Kim, P., Miller, S.A., Chu, T., Harris-White, M.E., Cole, G.M. *Phenolic anti-inflammatory antioxidant reversal of A β -induced cognitive deficits and neuropathology*. Neurobiol Aging 2001, 22: 993-1005.
27. Gasparini, L., Ongini, E., Wenk, G. *Non-steroidal anti-inflammatory drugs (NSAIDs) in Alzheimer's disease: Old and new mechanisms of action*. J Neurochem 2004, 91:521-36.
28. Eriksen, J.L., Sagi, S.A., Smith, T.E. et al. *NSAIDs and enantiomers of flurbiprofen target γ -secretase and lower A β 42 in vivo*. J Clin Invest 2003, 112: 440-9.
29. Morihara, T., Chu, T., Ubeda, O., Beech, W., Cole, G.M. *Selective inhibition of A β 42 production by NSAID R-enantiomers*. J Neurochem 2002, 83: 1009-12.
30. Yamamoto, S. *Mammalian lipoxygenases: Molecular structures and functions*. Biochim Biophys Acta 1992, 1128: 117-31.
31. Funk, C.D. *The molecular biology of mammalian lipoxygenases and the quest for eicosanoid functions using lipoxygenase-deficient mice*. Biochim Biophys Acta 1996, 1304: 65-84.
32. Manev, H., Uz, T., Sugaya, K., Qu, T. *Putative role of neuronal 5-lipoxygenase in an aging brain*. FASEB J 2000, 14: 1464-9.
33. Uz, T., Pesold, C., Longone, P., Manev, H. *Aging associated up-regulation of neuronal 5-lipoxygenase expression: Putative role in neuronal vulnerability*. FASEB J 1998, 12: 439-49.
34. Bigby, T.D. *The yin and yang of 5-lipoxygenase pathway activation*. Mol Pharmacol 2002, 62: 200-2.
35. Whitman, S., Gezgin, M., Timmermann, B.N., Holman, T.R. *Structure-activity relationship studies of nordihydroguaiaretic acid inhibitors toward soybean 12-human and 15-human lipoxygenase*. J Med Chem 2002, 45: 2659-66.
36. Haeggstrom, J.Z., Wetterholm, A. *Enzymes and receptors in the leukotriene cascade*. Cell Mol Life Sci 2002, 59: 742-53.
37. Funk, C.D. *Prostaglandins and leukotrienes: Advances in eicosanoid biology*. Science 2001, 294: 1871-5.
38. Werz, O., Klemm, J., Samuelsson, B., Radmark, O. *5-Lipoxygenase is phosphorylated by p38 kinase-dependent MAPK/AP kinases*. Proc Natl Acad Sci USA 2000, 97: 5261-6.
39. Salari, H., Braquet, P., Borgeat, P. *Comparative effects of indomethacin, acetylenic acids, 15-HETE, nordihydroguaiaretic acid, and BW755C on the metabolism of arachidonic acid in human leukocytes and platelets*. Prostaglandins Leukot Med 1984, 13: 53-60.
40. Kemal, C. *Reductive inactivation of soybean lipoxygenase-1 by catechols*. Biochemistry 1987, 26: 7064-72.
41. Skrzypczak-Jankun, E.E., McCabe, N.P., Selman, S.H., Jankun, J. *Curcumin inhibits lipoxygenase by binding to its central cavity: Theoretical and X-ray evidence*. Int. J. Mol. Med 2000, 6: 521-6.
42. Lim, G.P., Chu, T., Yang, F., Beech, W., Frautschy, S.A., Cole, G.M. *The curry spice curcumin reduces oxidative damage and amyloid pathology in an Alzheimer transgenic mouse*. J Neurosci 2001, 21: 8370-7.
43. McGeer, P.L., McGeer, E.G. *Inflammation, autotoxicity, and Alzheimer disease*. Neurobiol Aging 2001, 22: 799-809.
44. Mhatre, M.C., Floyd R.A., Hensley K. *Oxidative stress and neuroinflammation in Alzheimer's disease and amyotrophic lateral sclerosis: Common links and potential therapeutic targets*. J Alzheimer's Dis 2004, 6: 147-57.
45. West, M.S., Mhatre, M.C., Ceballos, A. et al. *The arachidonic acid 5-lipoxygenase inhibitor nordihydroguaiaretic acid inhibits TNF α activation of microglia and extends survival of G93A-SOD1 transgenic mice*. J Neurochem 2004, 91:133-43.
46. Frautschy, S.A., Hu, W., Kim, P., Miller, S.A., Chu, T., Harris-White, M.E., Cole, G.M. *Phenolic anti-inflammatory antioxidant reversal of A β -induced cognitive deficits and neuropathology*. Neurobiol Aging 2001, 22: 993-1005.

47. Ono, K., Hasegawa, K., Yoshiike, Y., Takashima, A., Yamada, M., Naiki, H. *Nordihydroguaiaretic acid potently breaks down pre-formed Alzheimer's beta-amyloid fibrils in vitro*. J Neurochem 2002, 81: 434-40.
48. Ono, K., Hasegawa, K., Naiki, H., Yamada, M. *Curcumin has potent anti-amyloidogenic effects for Alzheimer's beta-amyloid fibrils in vitro*. J Neurosci Res 2004, 75: 742-50.
49. Moss, M.A., Varvel, N.H., Nichols, M.R., Reed, D.K., Rosenberry, T.L. *Nordihydroguaiaretic acid does not disaggregate beta-amyloid(1-40) protofibrils but does inhibit growth arising from direct protofibril association*. Mol Pharmacol 2004, 66: 592-600.
50. Goodman, Y., Steiner, M.R., Steiner, S.M., Mattson, M.P. *Nordihydroguaiaretic acid protects hippocampal neurons against amyloid beta-peptide toxicity, and attenuates free radical and calcium accumulation*. Brain Res 1994, 654:171-6.
51. In, K.H., Asano, K., Beier, D. et al. *Naturally occurring mutations in the human 5-lipoxygenase gene promoter that modify transcription factor binding and reporter gene transcription*. J Clin Invest 1997, 99: 1130-7.
52. Qu, T., Manev, R., Manev, H. *5-Lipoxygenase (5-LOX) promoter polymorphism in patients with early-onset and late-onset Alzheimer's disease*. J Neuropsychiatry Clin Neurosci 2001, 13: 304-5.
53. Hada, T., Hagiya, H., Suzuki, H. et al. *Arachidonate 12-lipoxygenase of rat pineal glands: Catalytic properties and primary structure deduced from its cDNA*. Biochim Biophys Acta. 1994, 1211: 221-8.
54. Pratico, D., Zhukareva, V., Yao, Y. et al. *12/15-Lipoxygenase is increased in Alzheimer's disease: Possible involvement in brain oxidative stress*. Am J Pathol 2004, 164: 1655-62.
55. Woo, C.H., Eom, Y.W., Yoo, M.H. et al. *Tumor necrosis factor alpha generates reactive oxygen species via a cytosolic phospholipase A₂-linked cascade*. J Biol Chem 2000, 275: 32357-62.
56. Hensley, K., Fedynyshyn, J., Ferrell, S. et al. *Message and protein-level elevation of tumor necrosis factor alpha (TNF α) and TNF-modulating cytokines in spinal cords of the G93A-SOD1 mouse model for amyotrophic lateral sclerosis*. Neurobiol Disease 2003, 14: 74-80.
57. Perry, R.T., Collins, J.S., Wiener, H., Acton, R., Go, R.C. *The role of TNF and its receptors in Alzheimer's disease*. Neurobiol Aging. 2001, 22: 873-83.
58. Floden, A.M., Li, S., Combs, C.K. *Beta-amyloid-stimulated microglia induce neuron death via synergistic stimulation of tumor necrosis factor alpha and NMDA receptors*. J Neurosci 2005, 25: 2566-75.
59. Hensley, K., Maidt, M.L., Yu, Z., Sang, H., Markesbery, W.R., Floyd, R.A. *Electrochemical analysis of protein nitrotyrosine and dityrosine in the Alzheimer brain indicates region-specific accumulation*. J Neurosci 1998, 18: 8126-32.
60. Smith, M.A., Richey Harris, P.L., Sayre, L.M., Beckman, J.S., Perry, G. *Widespread peroxynitrite-mediated damage in Alzheimer's disease*. J Neurosci 1997, 17: 2653-7.
61. Su, J.H., Deng, G., Cotman, C.W. *Neuronal DNA damage precedes tangle formation and is associated with up-regulation of nitrotyrosine in Alzheimer's disease brain*. Brain Res 1997, 774: 193-9.
62. Funk, C.D. *Prostaglandins and leukotrienes: Advances in eicosanoid biology*. Science 2001, 294: 1871-5.
63. Brennan, P., O'Neill, L.A. *Inhibition of nuclear factor κ B by direct modification in whole cells – mechanism of action of nordihydroguaiaritic acid, curcumin and thiol modifiers*. Biochem Pharmacol 1998, 55: 965-73.
64. Bharti, A.C., Donato, N., Singh, S., Aggarwal B.B. *Curcumin (diferuloylmethane) down-regulates the constitutive activation of nuclear factor- κ B and I κ B α kinase in human multiple myeloma cells, leading to suppression of proliferation and induction of apoptosis*. Blood 2003, 101:1053-62.
65. Kuner, P., Schubnel, R., Hertel, C. *Beta-amyloid binds to p75NTR and activates NF κ B in human neuroblastoma cells*. J Neurosci Res 1998, 54: 798-804.
66. Dash, P.K., Moore, A.N. *Enhanced processing of APP induced by IL-1 β can be reduced by indomethacin and nordihydroguaiaretic acid*. Biochem Biophys Res Commun 1995, 208: 542-8.